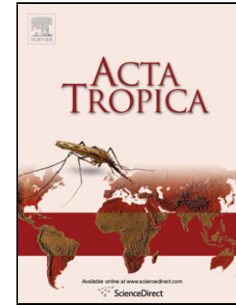


## Accepted Manuscript

Title: In Vitro Effects of Amino Alcohols on *Echinococcus granulosus*

Authors: Congshan Liu, Jianhai Yin, Jian Xue, Yi Tao, Wei Hu, Haobing Zhang



PII: S0001-706X(17)30811-2  
DOI: <http://dx.doi.org/10.1016/j.actatropica.2017.08.031>  
Reference: ACTROP 4423

To appear in: *Acta Tropica*

Received date: 6-7-2017  
Revised date: 21-8-2017  
Accepted date: 26-8-2017

Please cite this article as: Liu, Congshan, Yin, Jianhai, Xue, Jian, Tao, Yi, Hu, Wei, Zhang, Haobing, In Vitro Effects of Amino Alcohols on *Echinococcus granulosus*. *Acta Tropica* <http://dx.doi.org/10.1016/j.actatropica.2017.08.031>

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

**Short Communication*****In Vitro* Effects of Amino Alcohols on *Echinococcus granulosus***

Congshan Liu<sup>1,a</sup>, Jianhai Yin<sup>1, a</sup>, Jian Xue<sup>a</sup>, Yi Tao<sup>a</sup>, Wei Hu<sup>a,b</sup>, Haobing Zhang<sup>a,\*</sup>

<sup>a</sup> National Institute of Parasitic Diseases, Chinese Center for Disease Control and Prevention; Key Laboratory of Parasite and Vector Biology, MOH; National Center for International Research on Tropical Diseases; WHO Collaborating Centre for Tropical Diseases, Shanghai 200025, China

<sup>b</sup> Department of Microbiology and Microbial Engineering, School of Life Sciences, Fudan University, Shanghai 200433, China

\*Corresponding author: zhang\_haobing@163.com.

<sup>1</sup> There are joint first author.

**ABSTRACT**

Cystic echinococcosis is a globally distributed zoonotic disease, which is caused by the larval stage of *Echinococcus granulosus sensu lato*. The chemotherapy of the disease is limited to the use of benzimidazoles. Recently, mefloquine and its analogues, aminoalcohol-carbazole, and some amino alcohol derivatives were reported to display inhibitory effects on parasites. Here, the activities of 130 amino alcohol compounds against *E. granulosus* were tested on protoscoleces and germinal cells at a concentration of 20 µg/ml over a period of three days. As a result, sixteen compounds totally were effective against both protoscoleces and germinal cells, and their IC<sub>50</sub> and LC<sub>50</sub> were also calculated respectively. Then effects of the most active compounds were observed on metacestodes over 14 days *in vitro*. Although the structure of active compounds were variable, hydroxyl and amino groups connected by two carbon atoms are held in common as the key feature of these compounds. The further investigation on metacestodes incubated with these active compounds revealed that the effects of JF16 and BTB4 were comparable to that of mefloquine and mebendazole. In addition, the ultrastructure alternations induced by these compounds on *E. granulosus* were confirmed by scanning electron microscopy and transmission electron microscopy observations. In conclusion, amino alcohols were a class of compounds with efficacy against *E. granulosus*. The most effective compounds JF16 and

BTB4 indicated that their basic structure would be useful in the synthesis of new compound for the treatment of echinococcosis. However, their *in vivo* efficacy and toxicity need to be carefully evaluated in the future.

**Keywords:** Amino alcohols; *Echinococcus granulosus*; Protoscoleces; Germinal cells; Metacestodes.

## 1. Introduction

Cystic echinococcosis (CE) caused by larva stage of *Echinococcus granulosus sensu lato* is globally distributed. This disease affects humans and animals, and is very important in the aspect of health and economy (Brunetti *et al.*, 2010; Moro and Schantz, 2009). For most of patients, CE is always asymptomatic for many years until the parasites grow to an extent that triggers clinical signs, which can cause serious morbidity and death. And patients' life quality significantly reduces though treatment options such as surgery and chemotherapy that is necessary when patients with multiple cysts in two or more organs that not suitable for surgical removal, or is used as prophylaxis to secondary echinococcosis after surgery (Pawlowski *et al.*, 2001). In clinics, mebendazole and albendazole are the only drugs recommended by WHO for the treatment of cystic echinococcosis. However, both of these benzimidazoles are poorly absorbed by patients, resulting in the cure rate only about 30% (Moro and Schantz, 2009). For good clinical efficacy, the alternatives are needed.

Mefloquine, developed in 1971, is a synthetic analogue of quinine and is commonly used in malaria prophylaxis and treatment of chloroquine-resistant falciparum malaria (Leed *et al.*, 2002). It also has promising inhibitory activities against other parasites (Van Nassauw *et al.*, 2008; Walter *et al.*, 1987). Recently, mefloquine was shown to be effective against *E. multilocularis metacestodes* (Kuster *et al.*, 2011) and led to the death of *E. granulosus* protoscoleces and germinal cells *in vitro* (Liu *et al.*, 2015). Furthermore, several authors have reported the antiparasitic activity of different mefloquine analogues with amino group and alcohols group. In a previous study, an aminoalcohol-carbazole series with antimalarial properties was reported and a compound showed good efficacy in *P. burgher* when orally administrated in the mouse model (Molette *et al.*, 2013). And amino alcohol derivatives also displayed inhibitory effects on promastigote forms of *Leishmania chagasi* and *Leishmania amazonensis* (Coimbra *et al.*, 2010; Del Olmo *et al.*, 2002), *Trichomonas vaginalis* trophozoites (Giordani *et al.*, 2009), and trypomastigotes forms of *Trypanosoma cruzi* (Junior *et al.*, 2010). Moreover, Fernandes Fde

S *et al* reported the *in vitro* schistosomicidal activity of amino alcohol compounds (Fernandes Fde *et al.*, 2013).

Regarding the structure characterization and anti-parasitic activity of these aforementioned compounds, 130 selected amino alcohols' effects were tested on *E. granulosus in vitro* in the hope to find the potential active compounds as the indication for treatment of CE.

## 2. Materials and Methods

### 2.1. Chemicals and reagents

A total of 130 Amino-alcohol compounds were purchased from Maybridge (UK) with purity >90% and mefloquine hydrochloride was provided by Libang Pharmaceutical Co., Ltd. (Xi'an, China). All the compounds were dissolved in DMSO at 10 mg/ml as a stock solution. Dilutions of the stocking solution to the working concentration were prepared freshly with DMSO on the day of treatment according to the test concentration. The DMSO concentration was adjusted to 0.5% in all samples, which had no significant effects on treated parasites and cells. All culture media and reagents were purchased from Gibco-BRL (Zurich, Switzerland) while mebendazole and other reagents were from Sigma (St. Louis, MO, USA).

### 2.2 Parasites, animals and infection

Protoscoleces were collected from hydatid cysts in fresh sheep liver and rinsed 5–8 times with Hanks' Balanced Salt Solution containing penicillin G (500 U/ml) and streptomycin (500 U/ml) under sterile conditions, and protoscoleces with viability higher than 95% were used in the experiments. About 2,000 protoscoleces were inoculated intraperitoneally into each Kunming strain mouse which were purchased from SLAC Laboratory Animal Center (Shanghai, China). The genotypes of protoscoleces from sheep and germinal cells from secondary infected mice were reported to be G1 strain (Liu, 2015).

### 2.3. Ethics statement

Animal care and all animal procedures were carried out in compliance with the Guidelines for the Care and Use of Laboratory Animals produced by the Shanghai Veterinary Research Institute. The study was approved by the Ethics Committee of the National Institute of Parasitic Diseases, Chinese Center for Disease Control and Prevention. The license number was IPD-2014-2.

#### 2.4. *In vitro* culture of parasites

The rinsed protoscoleces were maintained as previously described (Liu, 2015) with minor modification. In brief, protoscoleces were cultured in RPMI 1640 medium supplemented with 10 % FBS, 10 % hydatid fluid, reducing agents ( $1 \times 10^{-5}$  M L-Dithiothreitol and 100  $\mu$ M L-cysteine), 2 mM glutamine, 1 mM sodium pyruvate, 100 U/ml penicillin G and 100  $\mu$ g/ml streptomycin at 37 °C in a 5 % CO<sub>2</sub> atmosphere. The culture medium was changed every 4-5 days. Mice secondary infected with *E. granulosus* for more than 10 months were sacrificed by cervical dislocation, and the cysts were carefully removed from the cavity of mice. After being washed several times with sterile PBS, the cysts with diameter <1 cm were cultured as protoscoleces. Then the germinal cells were prepared from larger cysts and cultured as previously described (Liu, 2015).

#### 2.5. *In vitro* drug treatment of parasites

The drug treatment *in vitro* was carried out in 96-well microtiter plates (Costar, USA) for protoscoleces and germinal cells. In this study, all the compounds were initially tested at 20  $\mu$ g/ml and the active ones were further studied at 0-20  $\mu$ g/ml. After cultivating for 1 week, protoscoleces were seeded at about 50 per well and germinal cells were plated with a density of about  $5 \times 10^4$  cells/ml per well in 200  $\mu$ l medium. Then the solution of compounds was added to the each well, using mefloquine and mebendazole as positive controls and RPMI 1640 medium and RPMI 1640 with 0.5 % DMSO as negative controls. Treatments were carried out at 37 °C in a 5 % CO<sub>2</sub> atmosphere for 72 h. Methylene blue exclusion method was used to test the viability of protoscoleces, in brief, 100  $\mu$ l 0.1% methylene blue was added to each well and observed under inverted microscope after 2 mins. The dead protoscoleces were stained blue and the surviving ones remained colorless. The viability of the germinal cells was determined by Cell Counting Kit-8 (Dojindo, Japan). The efficacy of active amino alcohols on metacestodes was carried out in 6-well microtiter plates (Costar, USA). Six metacestodes per well were collected with 6 ml medium. The tested final concentrations were 1, 5 and 10  $\mu$ g/ml and metacestodes were observed for 14 days. In parallel, germinal cells treated by active compounds at 10  $\mu$ g/ml were viewed by scanning electron microscopy (SEM), and the treated metacestodes were observed by SEM and transmission electron microscopy (TEM) to morphologically assess potential drug-induced damages. All experiments were carried out in triplicate and repeated at least twice.

## 2.6. Cytotoxicity test

Cytotoxicity was determined using three noncarcinogenic cells, L929 cells in RPMI1640, HK-2 cells in DMEM/F12 and Chang liver cells in DMEM medium, with 10% FBS; and 3 cancer cell lines, A172 cells in DMEM, A2058 cells in RPMI1640 and HCT-8 cells in DMEM medium, with 10% FBS. These cells were seeded in 96-well microtiter plates at a cell density of  $5 \times 10^4$  cells/well and allowed to attach for 24 h before treatment. Drug was added, and plates were incubated for 72 h. Cell viability was also determined by CCK-8 kit.

## 2.7. Ultrastructural investigations of drug-treated parasites

Samples of germinal cells and metacystodes treated by amino-alcohol compounds were processed for scanning electron microscopy and transmission electron microscopy (SEM and TEM), respectively. For investigating the ultrastructure of germinal cells, these cells were grown on glass coverslips before the drug treatment. Then freshly harvested parasites were fixed overnight at 4 °C in 2.5 glutaraldehyde in 0.1 M PBS (pH 7.4) and were post-fixed for more than 2h at 4 °C in 2% osmium tetroxide. Then these samples were proceeded for SEM and TEM observation (Liu *et al.*, 2016).

## 2.8. Data analysis

The values of LC<sub>50</sub> (the concentration that induced 50% protoscolecis death), IC<sub>50</sub> (the concentration that inhibited 50% germinal cell grow) and Tox<sub>50</sub> (the concentration that induced 50% cell toxicity) were calculated by probability unit method with SPSS version 17.0.

# 3. Results

## 3.1. Effectivities of 130 amino-alcohol compounds against protoscolecis and germinal cells in vitro

Firstly, A library of 130 amino alcohol compounds was screened against *E. granulosus* protoscolecis and germinal cells at 20 µg/ml over 3 days. Then the effective compounds were selected to determine the values of IC<sub>50</sub> and LC<sub>50</sub> (Table 1). Results from Table 1 showed that the active compounds had varied structures, but they had one thing in common that all of them contained hydroxyl group and amino group and these two groups were connected by two carbon atoms. In addition, the amino group in these compounds was nitrogen heterocyclic ring, such as carbazolyl, indole, piperidine and

piperazine. The replacement of trifluoromethyl moiety ( AW6 vs AW13) and removal of halogens (AW17 vs AW19) decreased activity on parasites. However, in carbozaole and benzhydrylpiperazino moiety, the removal of one or two halogens did not effect compounds' antiparasitic activity. Moreover, most of the amino-alcohol series showed better effects on germinal cells than protoscolecocytes at the same concentration, indicating the germinal cells was more sensitive to these compounds. For mefloquine and mebendazole, the values of calculated  $IC_{50}$  of germinal cells were very close to that of  $LC_{50}$  of protoscolecocytes (Table1).

### 3.2. The activity of selected active amino-alcohols against *E. granulosus* metacestodes *in vitro*

Compounds with prominent effects on *E. granulosus* protoscolecocytes and germinal cells *in vitro* were selected and observed the effects on cultured metacestodes. And diverse effects were induced by different compounds. After 14 days incubated with 0.5% DMSO, metacestodes did not show any morphological alterations (Table S1 in Supplementary File) and exhibited an intact germinal layer (Fig. 1). However, amino-alcohols at 10  $\mu\text{g/ml}$  resulted in morphological changes of the cysts, including the softening of the cysts and detachment of GL from LL. In addition, with the decline of chemical concentration, compounds of AW16, AW19, SC7, HT3 and HT24 had subtle effects on metacestodes, while other compounds (BTB3 (Liu, 2016), BTB4, JF16, MEF and MBZ) were still effective (Table S1 in Supplementary File).

### 3.3. Ultrastructural alterations of *E. granulosus* induced by effective compounds

Under the SEM, control germinal cells were round and adherent to the plastic culture matrix by abundant of cell substrate. During the incubation period with 0.5% DMSO, no ultrastructural alterations were observed (Fig. 2, a1). In contrast, the germinal cells showed dramatical damages after incubating with JF16, BTB4, MEF and MBZ at 10  $\mu\text{g/ml}$ . In general, all the compounds induced the detachment of cells from the culture matrix along with alternations of cell morphology, including the loss of cell integrity, the generation of cavities in the cell surface and cell shrinkage. Moreover, most of the cell adhesion morphological changed or disappeared after the *in vitro* treatment.

For all the tested compounds, the most significant alternation caused by these effective compounds was the detachment of the inner GL from the LL confirmed by SEM and TEM. Normally, GL contains different cell types and is the only tissue responsible for the proliferation of the parasites. The outer

surface of carbohydrate-rich LL stays close to host tissue and the inner surface connects to the GL. In the control treatment, the GL was closely connected to LL by numerous microtriches, and the cells in GL were arranged in order. In contrast, some or all of the GL were separated from the LL after being incubated with compounds and no normal cell could be found connecting to LL (Fig. 2, b2-b5). Under TEM, the GL of metacestodes treated with JF16 and MBZ became vacuolized and no intact cell could be found (Fig. 2, c2, c5). In addition, the microtriches were also morphological changed by these compounds (Fig. 2, c2-c5).

(a1-a5) SEM of germinal cells after incubation for 3 days; (b1-b2) SEM of metacestodes after incubation for 14 days. LL, Laminated layer; GL, germinal layer; (c1-c5) TEM of metacestodes after incubation for 14 days. Teg, tegument; v, vacuoles; Arrows point towards microtriches that protrude into the laminated layer.

#### 3.4. Cytotoxicity in noncarcinogenic mammalian cells and potential anti-cancer effects

The cytotoxicity of selected amino alcohols was assessed in mammalian cells using CCK-8 method. For three noncarcinogenic cells, L929, HK-2 and Chang liver, the  $Tox_{50}$  of all tested compounds were higher than 20  $\mu$ M, while the cancer cells (A172, A2058 and HCT-8) were more sensitive to these compounds with the values of  $Tox_{50}$  lower than 10  $\mu$ M (except HT24, HT3, SC7 and MBZ). The details can be seen in Table. S2 in Supplementary File.

## 4. Discussion

An appropriate screening method is crucial for high-throughput screening of drugs. In the present study, three models were used to evaluate the treatment effects of chemical compounds against *E. granulosus in vitro*. Among these models, protoscoleces and metacestodes are widely used in many anti-echinococcus drug studies (Albani and Elissondo, 2014; Verma *et al.*, 2014), but the reports about the use of *E. granulosus sensu lato* germinal cells were limited (Albani, 2010). As the most important cells in germinal layer, germinal cells are the only proliferative part of the parasite responsible for the formation of capsules or protoscoleces asexually (Eckert *et al.*, 2001), thus they are usually considered as the main target for chemotherapy. Based on the successful experiences of maintaining germinal cells from secondary infected cysts (Liu, 2015) and the use of stable cell counting method, germinal cells



were used for large-scale drug screening in this study. Compare to protoscoleces and metacystodes, the germinal cells are more suitable for the initial drug screening because of the abundant cells can be easily acquired from several infected mice and the cell activity can be rapidly and accurately determined by microplate reader. In contrast, so far, the activity of *E. granulosus* protoscoleces and metacystodes was still depend on the time-consuming manual judgment. In the present study, by the use of *E. granulosus* protoscoleces, germinal cells and cysts, some amino alcohols were indicated to have potential use in the finding of novel compounds for combating echinococcosis. For all the experiment, in order to absolutely dissolve the tested compounds, the final concentration of 0.5% DMSO was necessary, and the 0.5% DMSO was set as the negative control. Then the final results indicated the negative control did not influenced the activity of parasites during the observation period.

This study was inspired by the anthelmintic effects of mefloquine on *E. multilocularis* and *E. granulosus*. This antimalarial drug was initially discovered the promising antischistosomal properties (Van Nassauw, 2008; Zhang *et al.*, 2009) in mice and effects on *E. multilocularis in vitro* (Kuster, 2011), as well as its anthelmintic effects on *E. granulosus in vitro* in our previous study (Liu, 2015). After analyzing the structure characterization of this compound, the amino and hydroxyl groups were deduced to be the key factors responsible for its function. In order to verified our deduction, 130 compounds containing amino and alcohol groups were selected and purchased from the commercial chemical database, then tested effects on *E. granulosus* protoscoleces and germinal cells *in vitro*. The results showed that more than 20% compounds showed good effects (both of the percentage of dead protoscoleces and inhibition rate of germinal cells >80% ) on the parasites (data not shown), indicating the potential anti-echinococcus properties of amino-alcohols. Then LC<sub>50</sub> and IC<sub>50</sub> of these candidate compounds against *E. granulosus* protoscoleces and germinal cell *in vitro* were determined, respectively. Most of compounds shower more effective on germinal cells than protoscoleces, and some compounds' effects on germinal cells were comparable to MBZ and MEF. Then these compounds were further tested on *E. granulosus* cysts, which present the parasitic status of these parasites in host. Among the candidate amino alcohols, JF16, BTB3 and BTB4 with similar structures were proved to be the most effective compounds on metacystodes in the present study. These three compounds all contained aminoalcohol-carbazole structure, which could improve their membrane permeability to potentially increase the chances of the drug passing through membrane, including the plasma membrane and the LL (the key parasitic structure for the transportation of drug into the GL) of

the *E. granulosus* metacetodes. This hypothesis is consistent with their CLogP values (JF16, 5.43; BTB3, 6.47 and BTB4, 5.98). Furthermore, the structures of BTB4 and JF16 are very similar to the reported most promising compound (compound 12) against *Plasmodium falciparum* K1 (Molette, 2013), indicating the potential activity of the three compounds on other parasites.

Besides the efficacy against *E. granulosus*, these candidate amino alcohols were also shown to be active against tumor cells. It can be attributed to the common characteristics between malignant tumors and parasites (Koppenol *et al.*, 2011; Mukhopadhyay *et al.*, 2002; Pierce *et al.*, 2012), such as the intense cell division out of host control, the interaction with the host immune response, and the high level of metabolic activity dependent on aerobic glycolysis, etc. (Lancelot *et al.*, 2013). In addition, some anti-parasitic and anti-cancer agents are proved to be effective on both of *Echinococcus spp.* and cancer cells. As a classical antihelmintic drug, mebendazole showed prospective therapeutic effects for many tumors (Bodhinayake *et al.*, 2015; Larsen *et al.*, 2015). Mefloquine and mefloquine–oxazolidine derivatives are considered to be new useful antitumor agents with rational designs. Therefore, the effects of the candidate amino alcohols against cancer cell lines were also evaluated in the present study. And higher toxicities of these compounds for A172 cells, A2058 cells and HCT-8 cells than normal host cell lines were observed. These results re-confirmed the similarities of *E. granulosus* with cancer cells and indicated that these amino alcohol compounds could be used as prospective therapeutic agents for tumors.

In order to test the cytotoxicity of the candidate amino alcohols, their effects against mammalian cells were further evaluated *in vitro*. Most of them displayed the  $Tox_{50}$  of 25~50  $\mu$ M for three mammalian normal cells. These concentrations are 5-fold in excess of the measured  $IC_{50}$  values observed for germinal cells. This selectivity of activity indicated the amino alcohols' inherent specificity for *E. granulosus* germinal cells compared to mammalian cells. The safety profile of amino alcohols was also taken into account in many studies. The compound 12 was reported to be negative in micronucleus and AMES *in vitro* assay, but showed high inhibition of the hERG (Molette, 2013). Mefloquine inhibited the hERG  $K^+$  channels in a concentration and time-dependent manner (Lopez-Izquierdo *et al.*, 2011; Traebert *et al.*, 2004). Considering the toxicity of amino alcohols reported, the further study should put emphasis on structure modification to enhance the specificity of amino alcohols on parasites rather than host tissues. At the same time, the proper treatment regime for these compounds in the use for the treatment of echinococcosis needs to be carefully explored.

In conclusion, several amino alcohols with the hydroxyl group and amino group connected by carbon atoms were effective against *E. granulosus* protoscoleces and germinal cells. The further investigation on metacestodes showed that JF16 and BTB4 may exhibit their *in vitro* activity similar with MEF but not as good as MBZ. Amino alcohols showed prominent efficacy against *E. granulosus* *in vitro*, but their possible toxicity should be paid more attention. In the future, the active compounds' *in vivo* efficacy against experimentally infected animals, the bioavailability, pharmacokinetics, *in vivo* toxicity, and potential side effects need to be carefully evaluated.

#### **Financial support**

This research was supported by the National Natural Science Foundation of China (Grant no. 81401691 ) and General Program of Shanghai Municipal Commission of Health and Family Planning (Grant no. 201540170).

#### **Acknowledgments**

We thank Han Xiumin from Qinghai Provincial People's Hospital for the kind provision of *E. granulosus* protoscoleces.

#### **Appendix A. Supplementary data**

The following is Supplementary data to this article.

## Reference

- Albani CM, Elissondo MC, Cumino AC, Chisari A, Denegri GM, 2010. Primary cell culture of *Echinococcus granulosus* developed from the cystic germinal layer: biological and functional characterization. *Int J Parasitol.* 40, 1269-1275.
- Albani MC, Elissondo MC, 2014. Efficacy of albendazole in combination with thymol against *Echinococcus multilocularis* protoscoleces and metacestodes. *Acta Trop.* 140, 61-67.
- Bodhinayake I, Symons M, Boockvar JA, 2015. Repurposing mebendazole for the treatment of medulloblastoma. *Neurosurgery.* 76, N15-16.
- Brunetti E, Kern P, Vuitton DA, Writing Panel for the W-I, 2010. Expert consensus for the diagnosis and treatment of cystic and alveolar echinococcosis in humans. *Acta Trop.* 114, 1-16.
- Coimbra ES, de Almeida MV, Junior CO, Taveira AF, da Costa CF, de Almeida AC, Reis EF, da Silva AD, 2010. Synthesis and antileishmanial activity of lipidic amino alcohols. *Chem Biol Drug Des.* 75, 233-235.
- Combrinck JM, Mabothe TE, Ncokazi KK, Ambele MA, Taylor D, Smith PJ, Hoppe HC, Egan TJ, 2013. Insights into the role of heme in the mechanism of action of antimalarials. *ACS Chem Biol.* 8, 133-137.
- Davis A, Dixon H, Pawlowski ZS, 1989. Multicentre clinical trials of benzimidazole-carbamates in human cystic echinococcosis (phase 2). *Bull World Health Organ.* 67, 503-508.
- Davis A, Pawlowski ZS, Dixon H, 1986. Multicentre clinical trials of benzimidazolecarbamates in human echinococcosis. *Bull World Health Organ.* 64, 383-388.
- Del Olmo E, Alves M, Lopez JL, Inchausti A, Yaluff G, Rojas de Arias A, San Feliciano A, 2002. Leishmanicidal activity of some aliphatic diamines and amino-alcohols. *Bioorg Med Chem Lett.* 12, 659-662.
- Eckert J, Gemmell MA, Meslin FX. WHO/OIE Manual on Echinococcosis in Humans and Animals: a Public Health Problem of Global Concern. Paris: Office International des Epizooties; 2001.
- Fernandes Fde S, Rezende Junior CO, Fernandes TS, da Silveira LS, Rezende CA, De Almeida MV, de Paula RG, Rodrigues V, Da Silva Filho AA, Couri MR, 2013. Anthelmintic effects of alkylated diamines and amino alcohols against *Schistosoma mansoni*. *Biomed Res Int.* 2013, 783490.
- Giordani RB, De Almeida MV, Fernandes E, Franca da Costa C, De Carli GA, Tasca T, Zuanazzi JA, 2009. Anti-*Trichomonas vaginalis* activity of synthetic lipophilic diamine and amino alcohol derivatives.

Biomed Pharmacother. 63, 613-617.

Junior CO, Alves RO, Rezende CA, da Costa CF, Silva H, Le Hyaric M, Fontes AP, Alves RJ, Romanha AJ, de Almeida MV, 2010. Trypanocidal activity of lipophilic diamines and amino alcohols. *Biomed Pharmacother.* 64, 624-626.

Keiser J, Odermatt P, Tesana S, 2009. Dose-response relationships and tegumental surface alterations in *Opisthorchis viverrini* following treatment with mefloquine in vivo and in vitro. *Parasitol Res.* 105, 261-266.

Koppenol WH, Bounds PL, Dang CV, 2011. Otto Warburg's contributions to current concepts of cancer metabolism. *Nat Rev Cancer.* 11, 325-337.

Kuster T, Stadelmann B, Hermann C, Scholl S, Keiser J, Hemphill A, 2011. In vitro and in vivo efficacies of mefloquine-based treatment against alveolar echinococcosis. *Antimicrob Agents Chemother.* 55, 713-721.

Lancelot J, Caby S, Dubois-Abdesselem F, Vanderstraete M, Trolet J, Oliveira G, Bracher F, Jung M, Pierce RJ, 2013. *Schistosoma mansoni* Sirtuins: characterization and potential as chemotherapeutic targets. *PLoS Negl Trop Dis.* 7, e2428.

Larsen AR, Bai RY, Chung JH, Borodovsky A, Rudin CM, Riggins GJ, Bunz F, 2015. Repurposing the antihelminthic mebendazole as a hedgehog inhibitor. *Mol Cancer Ther.* 14, 3-13.

Leed A, DuBay K, Ursos LM, Sears D, De Dios AC, Roepe PD, 2002. Solution structures of antimalarial drug-heme complexes. *Biochemistry.* 41, 10245-10255.

Liu C, Zhang H, Yin J, Hu W, 2015. In vivo and in vitro efficacies of mebendazole, mefloquine and nitazoxanide against cyst echinococcosis. *Parasitol Res.* 114, 2213-2222.

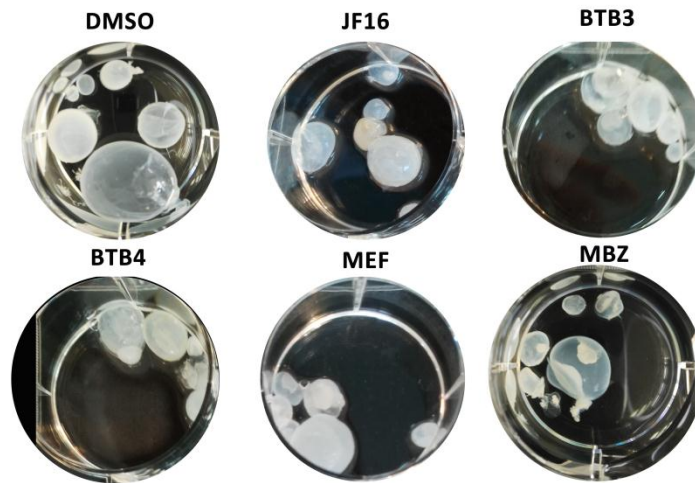
Liu CS, Zhang HB, Xue J, Tao Y, Hu W, 2016. In Vitro Effects of Aminoalcohol-carbazole Compound BTB3 against *Echinococcus granulosus*. *Zhongguo Ji Sheng Chong Xue Yu Ji Sheng Chong Bing Za Zhi.* 34, 245-248.

Lopez-Izquierdo A, Ponce-Balbuena D, Moreno-Galindo EG, Arechiga-Figueroa IA, Rodriguez-Martinez M, Ferrer T, Rodriguez-Menchaca AA, Sanchez-Chapula JA, 2011. The antimalarial drug mefloquine inhibits cardiac inward rectifier K<sup>+</sup> channels: evidence for interference in PIP<sub>2</sub>-channel interaction. *J Cardiovasc Pharmacol.* 57, 407-415.

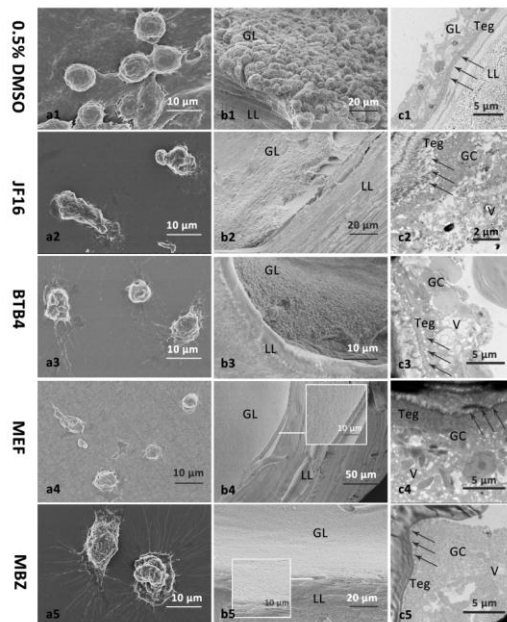
Mandal S, Mandal MD, 2012. Human cystic echinococcosis: epidemiologic, zoonotic, clinical, diagnostic and therapeutic aspects. *Asian Pac J Trop Med.* 5, 253-260.

- Manneck T, Keiser J, Muller J, 2012. Mefloquine interferes with glycolysis in schistosomula of *Schistosoma mansoni* via inhibition of enolase. *Parasitology*. 139, 497-505.
- Moazeni M, Saharkhiz MJ, Hosseini AA, 2012. In vitro lethal effect of ajowan (*Trachyspermum ammi* L.) essential oil on hydatid cyst protoscoleces. *Vet Parasitol*. 187, 203-208.
- Molette J, Routier J, Abia N, Besson D, Bombrun A, Brun R, Burt H, Georgi K, Kaiser M, Nwaka S, Muzerelle M, Scheer A, 2013. Identification and optimization of an aminoalcohol-carbazole series with antimalarial properties. *ACS Med Chem Lett*. 4, 1037-1041.
- Moro P, Schantz PM, 2009. Echinococcosis: a review. *Int J Infect Dis*. 13, 125-133.
- Mukhopadhyay T, Sasaki J, Ramesh R, Roth JA, 2002. Mebendazole elicits a potent antitumor effect on human cancer cell lines both in vitro and in vivo. *Clin Cancer Res*. 8, 2963-2969.
- Munkhjargal T, AbouLaila M, Terkawi MA, Sivakumar T, Ichikawa M, Davaasuren B, Nyamjargal T, Yokoyama N, Igarashi I, 2012. Inhibitory effects of pepstatin A and mefloquine on the growth of *Babesia* parasites. *Am J Trop Med Hyg*. 87, 681-688.
- Pawlowski ZS, Eckert J, Vuitton DA, Ammann RW, Kern P, Craig PS, Dar KF, De Rosa F, Filice C, Gottstein B, Grimm F, Macpherson CNL, Sato N, Todorov T, Uchino J, Von Sinner W, Wen H. Echinococcosis in humans: clinical aspects, diagnosis and treatment. In: Eckert J, Gemmell, M.A., Meslin, F.X., Pawlowski, Z.S., editor. WHO/OIE Manual on Echinococcosis in Humans and Animals: A Public Health Problem of Global Concern. Paris, France: World Organisation for Animal Health; 2001. p. 20-66.
- Pierce RJ, Dubois-Abdesselem F, Lancelot J, Andrade L, Oliveira G, 2012. Targeting schistosome histone modifying enzymes for drug development. *Curr Pharm Des*. 18, 3567-3578.
- Soukhathammavong P, Odermatt P, Sayasone S, Vonghachack Y, Vounatsou P, Hatz C, Akkhavong K, Keiser J, 2011. Efficacy and safety of mefloquine, artesunate, mefloquine-artesunate, tribendimidine, and praziquantel in patients with *Opisthorchis viverrini*: a randomised, exploratory, open-label, phase 2 trial. *Lancet Infect Dis*. 11, 110-118.
- Spicher M, Roethlisberger C, Lany C, Stadelmann B, Keiser J, Ortega-Mora LM, Gottstein B, Hemphill A, 2008. In vitro and in vivo treatments of echinococcus protoscoleces and metacestodes with artemisinin and artemisinin derivatives. *Antimicrob Agents Chemother*. 52, 3447-3450.
- Spiliotis M, Mizukami C, Oku Y, Kiss F, Brehm K, Gottstein B, 2010. Echinococcus multilocularis primary cells: improved isolation, small-scale cultivation and RNA interference. *Mol Biochem Parasitol*. 174, 83-87.

- Traebert M, Dumotier B, Meister L, Hoffmann P, Dominguez-Estevez M, Suter W, 2004. Inhibition of hERG K<sup>+</sup> currents by antimalarial drugs in stably transfected HEK293 cells. *Eur J Pharmacol.* 484, 41-48.
- Van Nassauw L, Toovey S, Van Op den Bosch J, Timmermans JP, Vercruysse J, 2008. Schistosomicidal activity of the antimalarial drug, mefloquine, in *Schistosoma mansoni*-infected mice. *Travel Med Infect Dis.* 6, 253-258.
- Verma VC, Gangwar M, Nath G, 2014. Osmoregulatory and tegumental ultrastructural damages to protoscoleces of hydatid cysts *Echinococcus granulosus* induced by fungal endophytes. *J Parasit Dis.* 38, 432-439.
- Verma VC, Gangwar M, Yashpal M, Nath G, 2013. Anticestodal activity of endophytic *Pestalotiopsis* sp. on protoscoleces of hydatid cyst *Echinococcus granulosus*. *Biomed Res Int.* 2013, 308515.
- Walter RD, Wittich RM, Kuhlow F, 1987. Filaricidal effect of mefloquine on adults and microfilariae of *Brugia patei* and *Brugia malayi*. *Trop Med Parasitol.* 38, 55-56.
- Xiao SH, Xue J, Li-li X, Zhang YN, Qiang HQ, 2010. Effectiveness of mefloquine against *Clonorchis sinensis* in rats and *Paragonimus westermani* in dogs. *Parasitol Res.* 107, 1391-1397.
- Xiao SH, Zhang CW, 2009. Histopathological alteration of juvenile *Schistosoma japonicum* in mice following treatment with single-dose mefloquine. *Parasitol Res.* 105, 1733-1740.
- Xue J, Jiang B, Liu CS, Sun J, Xiao SH, 2013. Comparative observation on inhibition of hemozoin formation and their in vitro and in vivo anti-schistosome activity displayed by 7 antimalarial drugs. *Zhongguo Ji Sheng Chong Xue Yu Ji Sheng Chong Bing Za Zhi.* 31, 161-169.
- Yamashita K, Uchino J, Sato N, Furuya K, Namieno T, 1997. Establishment of a primary culture of *Echinococcus multilocularis* germinal cells. *J Gastroenterol.* 32, 344-350.
- Zhang CW, Xiao SH, Utzinger J, Chollet J, Keiser J, Tanner M, 2009. Histopathological changes in adult *Schistosoma japonicum* harbored in mice treated with a single dose of mefloquine. *Parasitol Res.* 104, 1407-1416.



**Figure 1.** The morphology changes of *E. granulosus* murine metacestodes after incubation with 0.5% DMSO, JF16, BTB3, BTB4, MEF and MBZ at 10 µg/ml for 14 days *in vitro*



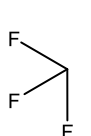
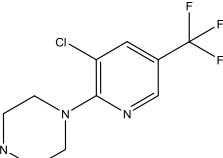
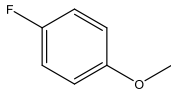
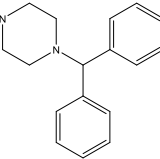
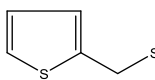
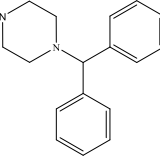
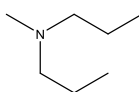
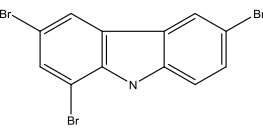
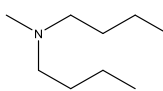
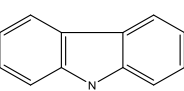
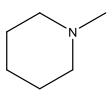
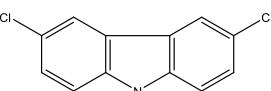
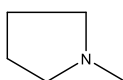
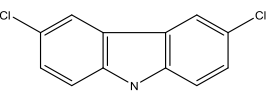
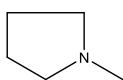
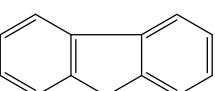
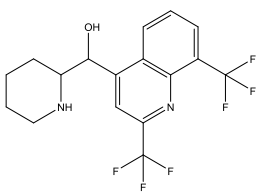
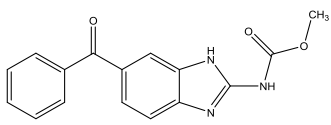
**Figure 2.** TEM and SEM observations of *E. granulosus* germinal cells and metacestodes incubated *in vitro* with active amino-alcohols at 10 µg/ml.



**Table 1.** *In vitro* effects of amino-alcohols against *E. granulosus* protoscolexes and germinal cells

OH  
R-CH-CH<sub>2</sub>-NHC

Compound	R	NHC	Protoscolexes, LC50, $\mu\text{M}$	Germinal cells, IC50, $\mu\text{M}$
AW6			18.38 $\pm 2.51$	11.34 $\pm 0.74$
AW13			16.09 $\pm 1.92$	5.18 $\pm 0.42$
AW16			11.55 $\pm 1.93$	4.81 $\pm 0.32$
AW17			21.70 $\pm 2.55$	5.45 $\pm 0.63$
AW19			14.08 $\pm 1.38$	3.01 $\pm 0.19$
HT10			21.04 $\pm 1.86$	6.11 $\pm 0.42$
HT20			17.59 $\pm 1.61$	12.23 $\pm 1.06$
HT22			21.54 $\pm 2.72$	5.37 $\pm 0.65$

SC7			11.70 ± 0.64	7.20 ± 0.82
HT3			8.97 ± 0.74	4.23 ± 0.52
HT24			6.43 ± 0.82	4.95 ± 0.33
BTB2			16.39 ± 1.70	4.40 ± 0.95
BTB3			12.82 ± 1.05	6.86 ± 1.25
BTB4			10.87 ± 1.48	6.28 ± 0.66
JF16			18.42 ± 1.79	5.04 ± 0.44
JF17			30.40 ± 3.36	20.04 ± 1.12
MEF			5.77 ± 1.19	4.47 ± 0.64
MBZ			7.88 ± 0.60	6.61 ± 0.70